

Two New Triterpenoids from the Stems of *Schisandra bicolor*

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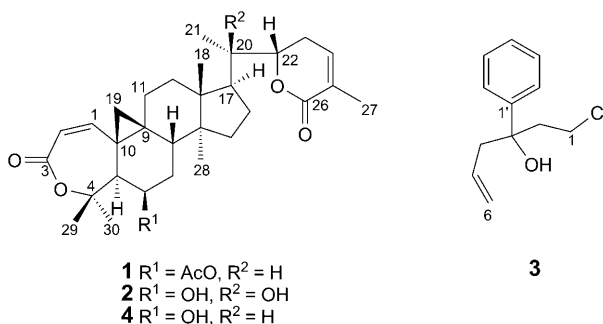
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Two new triterpenoids, schisanbilactones A and B (**1** and **2**, resp.), and a new natural product, 1-chloro-3-phenylhex-5-en-3-ol (**3**), as well as a known triterpenoid, kadsulactone A (**4**), were isolated from the stems of *Schisandra bicolor*. The structures of the new compounds were elucidated by spectroscopic analyses, including 2D-NMR techniques.

Introduction. – Plants of the family Schisandraceae have proved to be a rich source of dibenzocyclooctane lignans, as well as lanostane and cycloartane triterpenes, which have been found to possess many beneficial pharmacological effects, such as anti-lipid peroxidative, antitumor, and anti-HIV activities [1–5]. In our previous study, four new lignans and three new carotane sesquiterpenoids were isolated from the stems of *Kadsura induta* and *Schisandra wilsoniana*, and some of them were found to show an antiviral effect against the hepatitis B virus [6–8]. In the course of searching for new bioactive natural products from Schisandraceae plants, two new triterpenoids, schisanbilactones A and B (**1** and **2**, resp.), and a new natural product, 1-chloro-3-phenylhex-5-en-3-ol (**3**), as well as a known triterpenoid, kadsulactone A (**4**), were isolated from the stems of *Schisandra bicolor* CHENG, indigenous to southern China. This article describes the isolation and the structural elucidation of the new compounds.



Results and Discussion. – Repeated column chromatography of the Et₂O extract from the stems of *S. bicolor* yielded compounds **1–4**.

Schisanbilactone A (**1**) was obtained as a white powder and was shown to possess the molecular formula $C_{32}H_{44}O_6$ on the basis of HR-ESI-MS (m/z 547.3044 ($[M + Na]^+$)), which indicated eleven degrees of unsaturation. The IR spectrum indicated the presence of a six-membered α,β -unsaturated lactone (1718 cm^{-1}) and a seven-membered α,β -unsaturated lactone (1685 cm^{-1}) [9].

The $^1\text{H-NMR}$ spectrum (*Table 1*) showed a secondary Me group ($\delta(\text{H})$ 0.97, *d*, $J = 6.4$) and six tertiary Me groups ($\delta(\text{H})$ 0.92, 1.01, 1.41, 1.47, 1.90, and 2.03, *s*). As determined by DEPT experiment, the down-field signals corresponded to three CO groups ($\delta(\text{C})$ 166.5, 166.8, and 169.6), four olefinic ($\delta(\text{C})$ 119.8, 128.3, 139.4, and 150.7), and three O-bearing C-atoms ($\delta(\text{C})$ 70.5, 80.4, and 83.6); the shielded region showed seven Me ($\delta(\text{C})$ 13.1, 17.0, 18.0, 19.5, 21.2, 23.9, and 27.9), seven CH_2 ($\delta(\text{C})$ 23.4, 26.9, 28.5, 28.8, 32.4, 35.8, and 36.5), and four CH groups ($\delta(\text{C})$ 39.1, 40.7, 48.1, and 48.2), as well as four quaternary C-atoms ($\delta(\text{C})$ 27.9, 32.0, 45.6, and 47.7) (*Table 2*). The NMR spectra of **1** were very similar to those of kadsulactone A (**4**), suggesting that **1** has the same skeleton as **4**.

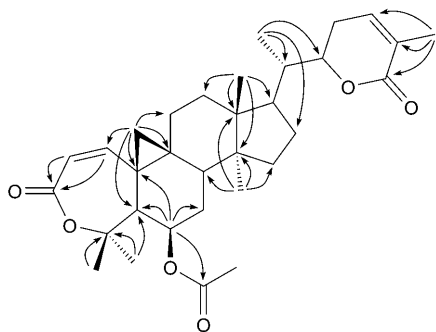
Table 1. $^1\text{H-NMR}$ Data of **1** and **2** (at 400 MHz, in CDCl_3 at 27° ; δ in ppm, J in Hz).

	1	2
H–C(1)	6.03 (<i>d</i> , $J = 12.7$)	6.03 (<i>d</i> , $J = 12.8$)
H–C(2)	5.90 (<i>d</i> , $J = 12.7$)	5.89 (<i>d</i> , $J = 12.8$)
H–C(5)	2.53 (<i>d</i> , $J = 3.6$)	2.32 (<i>d</i> , $J = 3.6$)
H–C(6)	5.27–5.33 (<i>m</i>)	4.44–4.48 (<i>m</i>)
CH_2 (7)	1.33–1.41 (<i>m</i>), 1.70–1.78 (<i>m</i>)	1.42–1.49 (<i>m</i>), 1.75–1.83 (<i>m</i>)
H–C(8)	1.77–1.85 (<i>m</i>)	2.18–2.26 (<i>m</i>)
CH_2 (11)	1.38–1.45 (<i>m</i>), 2.16–2.28 (<i>m</i>)	1.42–1.50 (<i>m</i>), 2.22–2.30 (<i>m</i>)
CH_2 (12)	1.67–1.80 (<i>m</i>)	1.78–1.86 (<i>m</i>)
CH_2 (15)	1.21–1.26 (<i>m</i>), 1.30–1.36 (<i>m</i>)	1.33–1.41 (<i>m</i>), 1.40–1.48 (<i>m</i>)
CH_2 (16)	1.40–1.53 (<i>m</i>), 1.75–1.84 (<i>m</i>)	1.55–1.63 (<i>m</i>), 1.96–2.03 (<i>m</i>)
H–C(17)	1.56–1.65 (<i>m</i>)	1.93–2.01 (<i>m</i>)
Me(18)	1.01 (<i>s</i>)	1.26 (<i>s</i>)
CH_2 (19)	1.23 (<i>d</i> , $J = 4.5$), 1.73 (<i>d</i> , $J = 4.5$)	1.17 (<i>d</i> , $J = 4.3$), 1.97 (<i>d</i> , $J = 4.3$)
H–C(20)	1.96–2.04 (<i>m</i>)	–
Me(21)	0.97 (<i>d</i> , $J = 6.4$)	1.32 (<i>s</i>)
H–C(22)	4.44 (<i>dq</i> , $J = 7.6, 13.2$)	4.29 (<i>dd</i> , $J = 4.5, 12.8$)
CH_2 (23)	1.95–2.07 (<i>m</i>)	2.26–2.33 (<i>m</i>)
H–C(24)	6.59 (<i>d</i> , $J = 6.4$)	6.60 (<i>d</i> , $J = 6.0$)
Me(27)	1.90 (<i>s</i>)	1.93 (<i>s</i>)
Me(28)	0.92 (<i>s</i>)	0.97 (<i>s</i>)
Me(29)	1.47 (<i>s</i>)	1.67 (<i>s</i>)
Me(30)	1.41 (<i>s</i>)	1.50 (<i>s</i>)
AcO	2.03 (<i>s</i>)	–

Comparison of the NMR spectra of **1** and **4** showed that the OH group at C(6) in **4** was replaced by an AcO group in **1**. The fragment ion at m/z 43 ($\text{C}_2\text{H}_3\text{O}^+$) in the EI-MS of **1** suggested the presence of an AcO group, as confirmed by the NMR signals at $\delta(\text{H})$ 2.03 (*s*), and $\delta(\text{C})$ 169.6 and 21.2. The HMBC correlations (*Fig. 1*) of $\delta(\text{H})$ 5.27–5.33 (H–C(6)) with $\delta(\text{C})$ 169.6 (C=O) revealed that the AcO group is located at C(6).

Table 2. ^{13}C -NMR Data of **1**, **2**, and **4** (at 100 MHz, in CDCl_3 at 27° ; δ in ppm)

	1	2	4
1	150.7 (<i>d</i>)	151.7 (<i>d</i>)	151.9 (<i>d</i>)
2	119.8 (<i>d</i>)	119.4 (<i>d</i>)	119.3 (<i>d</i>)
3	166.8 (<i>s</i>)	167.6 (<i>s</i>)	167.6 (<i>s</i>)
4	83.6 (<i>s</i>)	84.8 (<i>s</i>)	84.9 (<i>s</i>)
5	48.2 (<i>d</i>)	48.8 (<i>d</i>)	48.8 (<i>d</i>)
6	70.5 (<i>d</i>)	66.7 (<i>d</i>)	66.5 (<i>d</i>)
7	26.9 (<i>t</i>)	33.3 (<i>t</i>)	33.2 (<i>t</i>)
8	40.7 (<i>d</i>)	39.1 (<i>d</i>)	39.6 (<i>d</i>)
9	27.9 (<i>s</i>)	27.8 (<i>s</i>)	28.0 (<i>s</i>)
10	32.0 (<i>s</i>)	32.0 (<i>s</i>)	32.0 (<i>s</i>)
11	28.5 (<i>t</i>)	28.8 (<i>t</i>)	27.1 (<i>t</i>)
12	32.4 (<i>t</i>)	33.2 (<i>t</i>)	32.6 (<i>t</i>)
13	47.7 (<i>s</i>)	48.2 (<i>s</i>)	47.9 (<i>s</i>)
14	45.6 (<i>s</i>)	46.3 (<i>s</i>)	45.7 (<i>s</i>)
15	35.8 (<i>t</i>)	35.3 (<i>t</i>)	35.7 (<i>t</i>)
16	28.8 (<i>t</i>)	21.7 (<i>t</i>)	28.7 (<i>t</i>)
17	48.1 (<i>d</i>)	50.9 (<i>d</i>)	48.3 (<i>d</i>)
18	18.0 (<i>q</i>)	20.1 (<i>q</i>)	18.2 (<i>q</i>)
19	36.5 (<i>t</i>)	37.4 (<i>t</i>)	37.2 (<i>t</i>)
20	39.1 (<i>d</i>)	75.4 (<i>s</i>)	39.1 (<i>d</i>)
21	13.1 (<i>q</i>)	20.7 (<i>q</i>)	13.1 (<i>q</i>)
22	80.4 (<i>d</i>)	83.1 (<i>d</i>)	80.5 (<i>t</i>)
23	23.4 (<i>t</i>)	25.6 (<i>t</i>)	23.5 (<i>t</i>)
24	139.4 (<i>d</i>)	139.0 (<i>d</i>)	139.5 (<i>d</i>)
25	128.3 (<i>s</i>)	128.3 (<i>s</i>)	128.3 (<i>s</i>)
26	166.5 (<i>s</i>)	165.5 (<i>s</i>)	166.6 (<i>s</i>)
27	17.0 (<i>q</i>)	16.9 (<i>q</i>)	17.0 (<i>q</i>)
28	19.5 (<i>q</i>)	19.9 (<i>q</i>)	19.7 (<i>q</i>)
29	27.9 (<i>q</i>)	28.1 (<i>q</i>)	28.1 (<i>q</i>)
30	23.9 (<i>q</i>)	24.0 (<i>q</i>)	24.0 (<i>q</i>)
AcO	169.6 (<i>s</i>), 21.2 (<i>q</i>)	–	–

Fig. 1. Key HMBCs in **1**

Compound **1** was inferred to have (*S*)-configuration at C(22) on the basis of a negative *Cotton* effect at 240 nm and a positive *Cotton* effect at 281 nm, similar to those of **4** [9].

Schisanbilactone B (**2**), obtained as a white powder, had the molecular formula $C_{30}H_{42}O_6$ with ten degrees of unsaturation determined by HR-ESI-MS (m/z 521.2884 ($[M + Na]^+$)). The IR spectrum showed the presence of a OH group (3434 cm^{-1}). The NMR, IR, and UV spectra of **2** were very similar to those of kadsulactone A (**4**), indicating that **2** also has the same skeleton as **4**. The $^1\text{H-NMR}$ spectrum (Table 1) showed six tertiary Me groups ($\delta(\text{H})$ 0.97, 1.26, 1.32, 1.50, 1.67, and 1.93, 6s). Comparison of the $^1\text{H-NMR}$ spectra of **2** and **4** showed that the secondary Me signal present in the latter at $\delta(\text{H})$ 0.98 (*d*, $J=6.4$, Me(21)) had disappeared in **2**, instead appearing as a tertiary Me signal at $\delta(\text{H})$ 1.32. HMBCs between Me(21) at $\delta(\text{H})$ 1.32 and C(17) at $\delta(\text{C})$ 50.9, C(20) at $\delta(\text{C})$ 75.4, and C(22) at $\delta(\text{C})$ 83.1, indicated a OH group at C(20), similar as renchangelactone A [10]. The CD spectrum of **2** showed a negative Cotton effect at 237 nm and a positive effect at 281 nm, which indicated that **2** had (*S*)-configuration at C(22) similar to that of **1**, different from that of renchangelactone A [10].

Compound **3**, a colorless, optically active oil, had the molecular formula $C_{12}H_{15}ClO$ according to HR-EI-MS (m/z 169.0417 ($[M - \text{CH}_2\text{CH}=\text{CH}_2]^+$)). The IR spectrum showed the presence of a OH group (3448 cm^{-1}). The $^1\text{H-NMR}$ signals at $\delta(\text{H})$ 5.51–5.62 (*m*, 1 H), 5.18–5.20 (*m*, 2 H), and $^{13}\text{C-NMR}$ signals at $\delta(\text{C})$ 120.6 and 132.5, suggested a monosubstituted C=C bond in **3**. The fragment ion at m/z 41 and NMR signals, along with a HMBC (Fig. 2) of $\text{CH}_2(6)$ at $\delta(\text{H})$ 5.18–5.20 to C(4) at $\delta(\text{C})$ 47.8, indicated the presence of a $\text{CH}_2\text{CH}=\text{CH}_2$ group. The fragment ions at m/z 91 (C_7H_7^+), 77 (C_6H_5^+), and 51 (C_4H_3^+), as well as NMR signals (Table 3), suggested a monosubstituted Ph group in **3**. The HMBC between H–C(2') and H–C(6') and

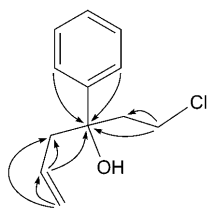


Fig. 2. Key HMBCs in **3**

Table 3. ^1H - and $^{13}\text{C-NMR}$ Data of **3** (at 400 and 100 MHz, resp., in CDCl_3 at 27° ; δ in ppm, J in Hz)

	$\delta(\text{H})$	$\delta(\text{C})$
$\text{CH}_2(1)$	3.20–3.29 (<i>m</i>), 3.56–3.65 (<i>m</i>)	40.2 (<i>t</i>)
$\text{CH}_2(2)$	2.32–2.38 (<i>m</i>)	45.5 (<i>t</i>)
C(3)	–	75.2 (<i>s</i>)
$\text{CH}_2(4)$	3.20–3.29 (<i>m</i>), 3.56–3.65 (<i>m</i>)	47.8 (<i>t</i>)
H–C(5)	5.51–5.62 (<i>m</i>)	132.5 (<i>d</i>)
$\text{CH}_2(6)$	5.18–5.20 (<i>m</i>)	120.6 (<i>t</i>)
C(1')	–	144.4 (<i>s</i>)
H–C(2',6')	7.36–7.45 (<i>m</i>)	128.5 (<i>d</i>)
H–C(3',5')	7.36–7.45 (<i>m</i>)	125.0 (<i>d</i>)
H–C(4')	7.27–7.33 (<i>m</i>)	127.0 (<i>d</i>)
OH	2.28 (<i>s</i>)	

C(3) at $\delta(C)$ 75.2 indicated that the Ph group was connected to C(3). Finally, the structure of **3** was elucidated as 1-chloro-3-phenylhex-5-en-3-ol of as yet unknown absolute configuration, based on the HSQC and HMBC spectra.

Experimental Part

General. Anal. TLC was performed on silica-gel plates (*Yan-tai Institute of Chemical Technology*), with petroleum ether (PE)/acetone 3:1 as eluent; visualization under UV light, and by spraying with 10% aq. H_2SO_4 , followed by heating. Column chromatography (CC): silica gel (SiO_2 ; 200–300, or 300–400 mesh; *Qingdao Marine Chemical Factory*). Optical rotations (ORD): *JASCO P-1020* spectropolarimeter. UV Spectra: *Shimadzu UV-260* spectrophotometer, in anh. MeOH; λ_{max} (log ϵ) in nm. CD Spectra: *JASCO J-715* spectropolarimeter; λ in nm ($\Delta\epsilon$). IR Spectra: *Avatar 360-ESP* spectrophotometer (*Thermo Nicolet*), as KBr pellets; in cm^{-1} . 1H - and ^{13}C -NMR Spectra: *DRX-400* spectrometer, in $CDCl_3$; δ in ppm, J in Hz. EI-MS: *Agilent 5973N* mass spectrometer; in m/z (rel. %). HR-ESI-MS: *Bruker APEX 70 TESLA FT-MS* apparatus. HR-EI-MS: *Waters Micromass GCT*.

Plant Material. The stems of *Schisandra bicolor* were collected at Tianmu mountain, Zhejiang Province, P. R. China, in June 2008. A voucher specimen (Qin-Ma200801) is deposited at the Herbarium of Materia Medica, School of Pharmacy, the Second Military Medical University, Shanghai, P. R. China.

Extraction and Isolation. The air-dried, ground stems (3 kg) of *S. bicolor* were extracted exhaustively with 95% aq. EtOH at r.t. The EtOH extract was concentrated *in vacuo* to yield a semi-solid (400 g), which was suspended in H_2O (400 ml), and extracted with Et_2O (3×400 ml). The combined org. phases were concentrated to yield a residue (100 g), part of which (95 g) was subjected to CC (2 kg of SiO_2 ; PE/AcOEt gradient) to afford seven fractions (*Frs. 1–7*). *Fr. 5*, eluted with PE/AcOEt 3:1, was subjected to repeated CC (SiO_2 ; PE/AcOEt 5:1), and then with PTLC (PE/AcOEt 5:3) to afford **1** (10 mg), **4** (36 mg), and **3** (10 mg). *Fr. 5*, eluted with PE/AcOEt 1:1, was subjected to repeated CC (SiO_2 ; PE/AcOEt 2:1), and then with PTLC (PE/acetone 3:2) to afford **2** (16 mg).

Schisanbilactone A (= (1*S*,3*aS*,5*R*,10*aS*,11*aS*,13*aR*)-1,2,3,3*a*,3*b*,4,5,5*a*,6,12,13,13*a*-Dodecahydro-3*a*,6,6,13*a*-tetramethyl-1-[(1*S*)-1-((2*S*)-5-methyl-6-oxo-3,6-dihydro-2H-pyran-2-yl)ethyl]-8-oxo-8H-cyclopenta[5,6]cyclopropa[1,8]naphtho[2,1-*c*]oxepin-5-yl Acetate; **1**). White powder. $[\alpha]_D^{25} = +72$ ($c = 0.05$, MeOH). UV (MeOH): 206 (4.18), 249 (4.26). CD ($c = 0.01$, MeOH): 240 (–4.5), 257 (–5.0), 281 (+32.2). IR (KBr): 2943, 1718, 1685, 1466, 1388, 1235, 1119, 1030. 1H - and ^{13}C -NMR: *Tables 1* and 2, resp. EI-MS: 446 (7), 169 (45), 107 (55), 105 (68), 95 (74), 93 (55), 91 (63), 55 (77), 43 (100). HR-ESI-MS: 547.3044 ($[M + Na]^+$, $C_{32}H_{44}NaO_6^+$; calc. 547.3031).

Schisanbilactone B (= (1*S*,3*aS*,5*R*,10*aS*,11*aS*,13*aR*)-1,2,3,3*a*,3*b*,4,5,5*a*,6,12,13,13*a*-Dodecahydro-5-hydroxy-1-[(1*R*)-1-hydroxy-1-((2*S*)-5-methyl-6-oxo-3,6-dihydro-2H-pyran-2-yl)ethyl]-3*a*,6,6,13*a*-tetramethyl-8H-cyclopenta[5,6]cyclopropa[1,8]naphtho[2,1-*c*]oxepin-8-one; **2**). White powder. $[\alpha]_D^{25} = +68$ ($c = 0.05$, MeOH). UV (MeOH): 206 (4.35), 252 (4.35). CD ($c = 0.01$, MeOH): 237 (–6.8), 281 (+28.1). IR (KBr): 3434, 2925, 1718, 1663, 1383, 1121, 915. 1H - and ^{13}C -NMR: *Tables 1* and 2, resp. EI-MS: 117 (19), 105 (12), 104 (100), 103 (50), 91 (79), 78 (40), 77 (24), 51 (22). ESI-MS: 415.1 ($[M + H]^+$). HR-ESI-MS: 521.2884 ($[M + Na]^+$, $C_{30}H_{42}NaO_6^+$; calc. 521.2874).

1-Chloro-3-phenylhex-5-en-3-ol (3). Colorless oil. $[\alpha]_D^{25} = +4.2$ ($c = 0.04$, MeOH). IR (KBr): 3448, 2926, 2366, 2345, 1654, 1638, 1447, 1060, 923, 702. 1H - and ^{13}C -NMR: *Table 3*. EI-MS: 171 (34), 170 (11), 169 (100), 105 (88), 93 (12), 91 (43), 77 (42), 63 (17), 41 (6). HR-EI-MS: 169.0417 ($[M - CH_2CH=CH_2]^+$, $C_9H_{10}ClO^+$; calc. 169.0420).

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